



Elsevier Editorial System(tm) for Food Hydrocolloids Manuscript Draft

Manuscript Number: FOODHYD-D-14-00640R1

Title: Wheat straw hemicellulose films as affected by citric acid

Article Type: Original Research Paper

Keywords: food packaging; polysaccharides; biopolymers; biodegradable films; crosslinking

Corresponding Author: Dr. Henriette M.C. de Azeredo,

Corresponding Author's Institution: Embrapa Agroindústria Tropical

First Author: Henriette M.C. de Azeredo

Order of Authors: Henriette M.C. de Azeredo; Charis Kontou-Vrettou, MSc; Graham K Moates, PhD; Nikolaus Wellner, PhD; Kathryn Cross, PhD; Paulo Henrique F Pereira; Keith W Waldron, PhD

Abstract: Hemicelluloses have been extracted from wheat straw with an alkaline peroxide solution. Biodegradable films intended for food packaging have been produced from hemicelluloses mixed with glycerol and different concentrations of citric acid (as a crosslinking agent), with or without sodium hypophosphite (catalyst to the crosslinking reaction). A curing treatment at 150°C has been carried out on the dried films in order to promote formation of ester bonds between citric acid and hemicelluloses. The crosslinking reaction, evidenced by FTIR spectra, improved water resistance and water vapor barrier properties of the films. On the other hand, the citric acid effects on film tensile properties were more consistent with those of a plasticizer than of a crosslinker, which might be ascribed to a flexible crosslinking. Sodium hypophosphite (SHP) did not affect the properties of crosslinked films. FTIR spectra indicated that some crosslinking reaction occurred even in a non-cured film.





1 Wheat Straw Hemicellulose Films as Affected by Citric Acid

- 2 Henriette M. C. Azeredo^{1,*}, Charis Kontou-Vrettou², Graham K. Moates³, Nikolaus
- 3 Wellner³, Kathryn Cross³, Paulo H.F. Pereira³, Keith W. Waldron³

4

¹Embrapa – Secretariat for International Affairs, Edifício Embrapa Sede, Prédio
CECAT, 3° andar, Parque Estação Biológica, Av. W3 Norte, Brasília, DF, Brazil,
CEP 70770-901, henriette.azeredo@embrapa.br; ²Agricultural University of Athens,
Iera Odos 75, 11855, Athens, Greece; ³Institute of Food Research, Norwich
Research Park, Colney, Norwich, NR4 7UA, UK.





10 Abstract

11 Hemicelluloses have been extracted from wheat straw with an alkaline peroxide 12 solution. Biodegradable films intended for food packaging have been produced from 13 hemicelluloses mixed with glycerol and different concentrations of citric acid (as a crosslinking agent), with or without sodium hypophosphite (catalyst to the 14 15 crosslinking reaction). A curing treatment at 150°C has been carried out on the dried 16 films in order to promote formation of ester bonds between citric acid and 17 hemicelluloses. The crosslinking reaction, evidenced by FTIR spectra, improved 18 water resistance and water vapor barrier properties of the films. On the other hand, 19 the citric acid effects on film tensile properties were more consistent with those of a 20 plasticizer than of a crosslinker, which might be ascribed to a flexible crosslinking. 21 Sodium hypophosphite (SHP) did not affect the properties of crosslinked films. FTIR 22 spectra indicated that some crosslinking reaction occurred even in a non-cured film.

23 Keywords: food packaging; polysaccharides; biopolymers; biodegradable films;
24 crosslinking.





25 **1. Introduction**

26 In the last decades, an increasing interest from research institutes and industries in 27 biopolymers from renewable sources is noticeable, and has been motivated by 28 shortage of natural energy sources as well as the necessity of replacement of 29 petroleum-based products, which is connected with demands for more environmentally friendly materials. In 2011, the global use of biodegradable plastics 30 31 was 0.85 million metric tons. BCC Research recorded a global bioplastic demand of 32 1.1 million metric tons in 2013, expected to reach 1.4 million metric tons in 2014 and about 6 million metric tons in 2019, in a compound annual growth rate of 32.7% 33 34 (BCC, 2014). According to studies by Helmut Kaiser Consultancy (2013), bioplastics 35 are expected to cover approximately 25-30% of the total plastics market by 2020.

36 The worldwide wheat consumption has been estimated to be around 705 million tons 37 in 2013/2014 (WASDE, 2014). Wheat straw and bran, which are abundant co-38 products of wheat crop, represent a valuable source of hemicelluloses, cellulose and lignin. In the European Union, some millions of tons of wheat straw and wheat bran 39 40 could be collected per year (Martel, Estrine, Plantier-Royon, Hoffmann, & Portella, 41 2010). Some of those co-products are used in animal feed and paper production, but 42 the majority is discarded as waste, when it could be exploited to produce high-valued 43 materials such as biodegradable (or edible) films for food packaging.

The hydrophilic nature of polysaccharide films provide them with good oxygen barrier properties, but their water vapor barrier and moisture resistance are poor (Giancone et al., 2011; Janjarasskul & Krochta, 2010). A high water solubility obviously affects





47 the applicability of edible films, since a packaging material is not supposed to 48 dissolve upon contact with water. Another problem which can result from polysaccharide hydrophilicity is swelling by water, leading the films to have their 49 50 mechanical and overall barrier properties impaired (Sebti, Delves-Broughton, & 51 Coma, 2003). The water resistance of a film can be improved by crosslinking, which 52 consists of linking polymer chains by covalent (chemical crosslinking) or by weaker 53 bonds (physical crosslinking), forming three-dimensional networks which reduce the mobility of the structure, usually enhancing its water resistance (reducing both water 54 55 solubility and swelling by water) as well as its mechanical and barrier properties 56 (Balaguer, Gómez-Estaca, Gavara, & Hernandez-Muñoz, 2011).

57 Dicarboxylic and polycarboxylic acids such as citric acid have been reported by several authors (Bonilla, Talón, Atarés, Vargas, & Chiralt, 2013; Coma, Sebti, 58 59 Pardon, Pichavant, & Deschamps, 2003; Möller, Grelier, Pardon, & Coma, 2004; 60 Olsson, Hedenqvist, Johansson, & Järnström, 2013a) to act as crosslinking agents 61 for polysaccharide films. One advantage of citric acid is that any unreacted acid is not only nutritionally acceptable but it can also act as a plasticizer (Chabrat, 62 63 Abdillahi, Rouilly, & Rigal, 2012; Shi et al., 2008). The crosslinking mechanism is 64 attributed to covalent intermolecular di-ester linkages between hydroxyl groups of the 65 polysaccharide and two carboxyl groups of the crosslinker (Coma et al., 2003; Hashem, Sharaf, El-Hady, & Hebeish, 2013; Olsson et al., 2013a), as indicated in 66 67 Figure 1. In a first step (not shown in Figure 1), carboxyl groups of the acid form 68 cyclic anhydrides, which react further with the carbohydrate hydroxyls (Xiaohong & Yang, 2000). The reaction has been reported to be favored by a high temperature 69 (typically above 100°C) curing process on the dried films (Coma et al., 2003; 70





71 Dastidar & Netravali, 2012; Olsson et al., 2013a, 2013b), although the crosslinking 72 reaction has also been reported to take place at a relatively low temperature (70°C) by Menzel et al. (2013). Sodium hypophosphite (SHP) acts as a catalyst for the 73 74 reaction (Reddy & Yang, 2010; Salam, Pawlak, Venditti, & El-tahlawy, 2011) by 75 weakening the hydrogen bonding between the carboxylic acid groups (Xiaohong & 76 Yang, 2000), increasing the speed of formation of the cyclic anhydride intermediate 77 (Feng, Xiao, Sui, Wang, & Xie, 2014; Garcia et al., 2014; Sauperl & Stana-78 Kleinschek, 2010) and allowing the reaction to occur at a lower temperature (Feng et 79 al., 2014). Peng, Yang, & Wang (2012), when studying maleic acid crosslinking in 80 cotton fabrics, suggested that SHP reacted to two maleic acid molecules already 81 esterified with cellulose, forming a new crosslink between two cellulose molecules.

A previous study (Ruiz et al., 2013) has shown wheat straw hemicelluloses as a
promising reinforcement material to κ-carrageenan/locust bean gum blend films.
However, no previous study has been found describing the use of wheat straw
hemicelluloses as a film matrix.

The objectives of this study were to evaluate some physical properties of films from wheat straw hemicelluloses as affected by citric acid contents, the presence of sodium hypophosphite and the curing step.

89 2. Materials and Methods

90 **2.1. Isolation of hemicelluloses**





91 Wheat straw was milled to less than 0.5 mm in a Retsch Brinkmann ZM-1 centrifugal 92 grinding mill (Retsch GmbH, Haan, Germany). The hemicellulose and α -cellulose 93 contents of milled wheat straw were analyzed (in triplicate) as described by TAPPI 94 T203cm-99 (TAPPI, 2009), and holocellulose, according to Yokoyama, Kadla, and 95 Chang (2002).

100 g of milled straw were washed with 1 L of 0.2% (w/v) ethylenediamine 96 97 tetraacetic acid (EDTA) solution at 90°C for 1 h, to remove water soluble 98 components and to chelate hydrogen peroxide (H_2O_2) decomposing metals. The 99 hemicelluloses were extracted with an alkaline H₂O₂ solution (2% w/v, pH adjusted to 100 12.5 with potassium hydroxide) for 16 h at 50°C, following the conditions as 101 optimized by Fang, Sun, Salisbury, Fowler, & Tomkinson (1999) for hemicellulose extraction of wheat straw. The H_2O_2 was used to delignify the material, since 102 103 preliminary tests indicated that films from non-delignified hemicelluloses (extracted 104 by using KOH solutions in different concentrations) were cracking, not continuous 105 structures. During an alkaline H₂O₂ treatment, the peroxide degrades in a reaction with the hydroperoxide anion (HOO⁻), producing the highly reactive hydroxyl radical 106 107 (HO⁻) which oxidizes lignin forming low molecular weight, water-soluble products 108 (Fang et al., 1999).

The supernatant resulting from the subsequent centrifugation was vacuum filtered and its pH was adjusted to 5.0 with acetic acid. Ethanol was added to a concentration of 60%, the suspension was stirred for 30 min, left overnight at 4°C, centrifuged, washed with ethanol 70%, centrifuged again, and left to dry at room





temperature. The dried hemicelluloses were then milled to a fine powder by using abasic mill (A10, IKA GmbH, Germany).

115 **2.2. Film formation**

116 The hemicellulose powder was homogenized for 15 min in distilled water (5 g/100 mL) with 30% glycerol (w/w, on a hemicellulose basis), citric acid (5%, 10%, 20%, or 117 118 30% w/w on a hemicellulose basis), with or without SHP (50% w/w on a citric acid 119 basis), by using a homogenizer (Ystral X10/25, Ballrechtar-Dottingen, Germany). Air 120 bubbles were removed under vacuum, and the films were cast on petri dishes to a final dried thickness of 0.09 mm, left to dry at room temperature for 24 h, then 121 122 conditioned (50% RH, 24°C) in an environmental chamber (Weiss Gallenkamp, 123 Loughborough, UK) for 24 h, so they had all similar moisture contents (between 124 11.91 and 12.32 g/100g). The dried films were then subjected to a curing treatment 125 at 150°C for 10 minutes using a fan oven (Memmert, Schwabach, Germany). Apart from a control film (with no citric acid or SHP), three groups of films were prepared, 126 127 namely, CA-C (those added with citric acid and cured), CA-NC (added with citric acid 128 and not subjected to the curing treatment), and CA-C-SHP (added with citric acid and SHP, and cured). 129

The water vapor permeability (WVP) determination was modified from the method E96-05 (ASTM, 2005) for five circular samples (30 mm in diameter). The thicknesses of the samples were measured using a micrometer screw gauge (Moore & Wright, Sheffield, UK) to the nearest 0.01 mm at 5 random locations, and the average value was calculated. The test films were sealed as patches onto acrylic permeation cells (2.4 cm in diameter and 1 cm in height) containing 2 mL of distilled water. The cells





were placed in a desiccator connected to two channels providing a steady flow of dried air (less than 1% RH) from a Balston 75-60 air drier at 24 °C, and were weighed 7 times over a 24-h period.

139 The water solubility determination was conducted on 2 cm x 2 cm film pieces in 140 quadruplicate, based on the method proposed by Ojagh, Rezaei, Razavi, & Hosseini 141 (2010), with some modifications. Previously dried and weighed samples were 142 immersed in 50 mL of distilled water for 6 h at 25°C, under stirring (150 rpm). The dry weight of the remaining film pieces was obtained after filtration on previously dried 143 144 and weighed filter paper, and it was used to calculate the insoluble matter as a 145 percentage of the initial dry weight. All the dry weights (of the initial and final film pieces and the filter paper) were determined after drying at 103°C for 24 h using a 146 147 fan oven (Memmert, Schwabach, Germany).

148 Tensile tests were conducted on 8 mm x 50 mm specimens, according to the method D882-09 (ASTM, 2009), on a Texture Analyzer TA.XT Plus (Stable Micro Systems, 149 Godalming, UK), using A/TG Tensile Grips and a 5 kg load cell, on 50 mm x 8 mm 150 151 strip film samples previously conditioned (50% RH, 24°C, 48 h) in an environmental chamber (Weiss Gallenkamp, Loughborough, UK). The thicknesses of the 152 specimens were determined by using a micrometer screw gauge (Moore & Wright, 153 154 Sheffield, UK) to the nearest 0.01 mm at 5 random locations. The initial grip separation and crosshead speed were set to 40 mm and 1 mm/s, respectively. Force 155 156 (N) and deformation (mm) were recorded during extension. Tensile strength was 157 calculated by dividing the required force for film rupture by the cross-sectional area, 158 and elongation at break was calculated as the percentage increase in sample length.





159 The elastic modulus was calculated from the slope of the stress-strain curve at the 160 elastic deformation region. The reported values correspond to five measurements.

Fourier-transform infrared (FTIR) spectra (128 scans at 2 cm⁻¹ resolution for a spectral range from 4000-500 cm⁻¹) were recorded using a Digilab FTIR spectrometer equipped with a Digilab UMA 600 microscope (Digilab, Randolph, MA, USA).

The scanning electron microscopy (SEM) images of gold-coated film surfaces were taken using a Zeiss Supra 55 VP SEM (Zeiss, Oberkochen, Germany) with an acceleration voltage of 3 kV, and a magnification of 1000 x.

168 **3. Results and Discussion**

169 The holocellulose, α-cellulose and hemicellulose contents of the wheat straw and the 170 hemicellulose powder are presented at Table 1. The contents of the wheat straw were similar to those reported by Govumoni, Koti, Kothagouni, Venkateshwar, & 171 Linga (2013). The hemicellulose powder (product of the extraction) contained mostly 172 hemicelluloses (about 87%), although there were still some α -cellulose and about 173 174 9% of other components. Considering that the yield of the extraction process was 18.5% w/w (on a wheat straw dry basis), and taking into account the hemicellulose 175 content of the wheat straw and the hemicellulose powder, the efficiency of the 176 extraction procedure can be estimated as only about 54%. 177

The film properties as affected by citric acid and SHP are presented in Figure 2. The most dramatic effect of citric acid was on insoluble matter, which was increased 5-





180 fold by the addition of 20% citric acid. The water vapor permeability (WVP) tended to 181 decrease accordingly, although not to the same extent. Some previous studies corroborate those changes (Coma et al., 2003; Menzel et al., 2013; Olivato, 182 183 Grossmann, Bilck, & Yamashita, 2012; Wang, Ren, Li, Sun, & Liu, 2014), which are attributed to the ester bonds formed between citric acid and the polysaccharide 184 (Wang et al., 2014), producing a denser structure. Crosslinking interconnects 185 polysaccharide molecules, providing better intermolecular interaction leading to a 186 decreased water uptake (Abdillahi, Chabrat, Rouilly, & Rigal, 2013). The reduced 187 188 polymer chain mobility probably made the water diffusion more difficult, explaining the reduced WVP (Olivato et al., 2012). The reduced availability of hydroxyl groups 189 190 and their substitution by hydrophobic ester groups could also contribute to the citric 191 acid content effects on WVP and water solubility (Ghanbarzadeh, Almasi, & Entezami, 2011; Ma, Jian, Chang, & Yu, 2008). 192

193 Some studies have reported citric acid effects on films only as a crosslinking agent (Olivato et al., 2012; Reddy & Yang, 2010; Reddy, Jiang, & Yang, 2012), while 194 195 others have reported it to act only as a plasticizer (Abdillahi et al., 2013; Chabrat et 196 al., 2012). However, the present study presented both effects apparently coexisting, although apparently inconsistent to each other. On the one hand, citric acid addition 197 decreased tensile strength and modulus, and increased elongation when at 10% 198 (and at higher concentrations for films with SHP), reflecting the behavior of a 199 plasticizer instead of a crosslinking agent. On the other hand, the water solubility and 200 201 the WVP tended to decrease with citric acid – the soluble matter increasing typically up to 20% citric acid, and the WVP tending to decrease up to 20% citric acid (except 202 for film with SHP, whose WVP did not significantly change). Some other studies 203





204 reported similar findings (Sebti et al., 2003; Shi et al., 2008; Wang et al., 2014). 205 Wang et al. (2014) observed that citric acid contents higher than 10% in polyvinyl alcohol/xylan films resulted in plasticizing effects evidenced from tensile tests, while 206 207 the water vapor permeability of the films kept decreasing with increasing citric acid contents, consistent with a crosslinking effect, suggesting coexisting crosslinking and 208 plasticizing effects of citric acid. The plasticizing effect of citric acid was attributed to 209 210 increasing interstitial volume of the film or increasing molecular mobility, making the 211 polymeric networks less dense (Wang et al., 2014). In the present study, although 212 citric acid impaired strength and modulus, it improved the toughness (which represents the energy required for the sample to break), corroborating results by 213 214 Reddy, Li, & Yang (2009). A possible explanation for this apparently inconsistent 215 behavior is that citric acid might act as a flexible crosslinker. Some previous studies 216 (Hu, Song, Liu, & Zhang, 2010; Jia, Zhang, He, & Ning, 2005; Zhang, Hu, Jia, & Du, 217 2003) have described that, although chemical crosslinking usually imposes 218 constraints on the motion of chain segments, a flexible crosslinking agent may act similarly to a plasticizer. The flexibility of citric acid crosslinking might then explain 219 why it was effective in reducing the film solubility and WVP while impairing the 220 221 overall tensile properties.

Although SHP has been mentioned as an effective catalyst for the crosslinking reaction of polysaccharides with citric acid (Feng et al., 2014; Reddy & Yang, 2010; Xiaohong & Yang, 2000), some studies (Coma et al., 2003; Feng et al., 2014) reported a lack of effect of SHP as a catalyst for citric acid crosslinking. On the other hand, the present study indicates that SHP had the opposite effect (especially at higher citric acid contents), decreasing strength and modulus, increasing elongation,





and increasing WVP, indicating that its presence actually impaired the crosslinking
effects of citric acid, which might be explained by its diluting effects on the reagents
(citric acid and hemicelluloses).

The non-cured films (CA-NC) presented tensile properties almost identical to those of the corresponding cured films (CA-C). Their only differences when compared to cured films were that their insoluble matter was lower, and, accordingly, their WVP was slightly higher. So, although the curing treatment has improved the water resistance of the crosslinked films, some crosslinking seems to have occurred even in non-cured films.

Figure 3 shows the Fourier transform infrared (FTIR) spectra of the control film (CA0) 237 and films with 30% citric acid (CA30-C, CA30-C-SHP, and CA30-NC). The peak at 238 1717 cm⁻¹ and the shoulder at 1695 cm⁻¹ are ascribed to free and hydrogen bonded 239 240 carboxylic acid groups, respectively. This area is less intense in CA30-C-SHP than in CA30-C film, indicating that SHP could have promoted citric acid decarboxylation on 241 curing, as suggested by Yao, Wang, Ye, & Yang (2013), which would explain the 242 unexpected effects of SHP on tensile properties and WVP of films in the present 243 study. The increased peak at 1208 cm⁻¹ in films with citric acid is ascribed to C-O 244 245 stretching of formed ester bonds (Bagheri, Yarmand, Madadlou, & Mousavi, 2014; Groen & Roberts, 2001; Rhim et al., 2004). The presence of ester bonds may be 246 indicated by the presence of a small shoulder around 1730 cm⁻¹ in films with citric 247 acid (Coma et al., 2003), which corresponds to ester C=O stretching vibration 248 249 (Dastidar & Netravali, 2012; Shi et al., 2007; Wang et al., 2014). Interestingly, this shoulder is evident even in the non-cured film, indicating again that some 250





251 crosslinking may have taken place even at room temperature, corroborating the 252 results of insoluble matter and WVP (Fig.2, E and F). Previously to this study, the lowest temperature at which the crosslinking reaction of citric acid with a 253 254 polysaccharide has been reported to take place was 70°C (Menzel et al., 2013). Moreover, the presence of SHP did not enhance this peak, suggesting that SHP did 255 not affect the crosslinking reaction, corroborating our previous observations on 256 257 physical properties of the films. The presence of SHP did not change noticeably the 258 spectra.

The curing process probably favoured not only crosslinking but also some degree of hydrolysis of the glycosidic bonds in hemicelluloses (especially with higher citric acid contents), which has been previously reported to occur in starch (Olivato et al., 2012; Shi et al., 2007; Menzel et al., 2013). However, the decreasing water solubility of the films with increasing citric acid contents (Fig. 2F) suggests that the hydrolysis effect was less noticeable than the crosslinking effect.

Glycerol (which was used as a plasticizer) can easily react with citric acid, competing with hemicelluloses, which could eventually impair the crosslinking. On the other hand, Yao et al. (2013) reported that glycerol and other polyols may act as crosslinking extenders, rather facilitating the crosslinking reaction. A study evaluating the effects of varying glycerol contents on crosslinked film properties would be worth investigating.

The SEM images (Figure 4) reveal that, while the control sample (C) was relatively smooth, the samples with citric acid (A and B) presented textured surfaces, corroborating Bonilla et al. (2013), who observed that the presence of citric acid 13





274 promoted a coarser surface on starch-chitosan films. The surface texture appears 275 like crystals (probably citric acid crystals) on the sample without SHP (A), but like ripples on the sample with SHP (B). The ripples may be due to the curing treatment, 276 277 and they may have been masked by the crystals on the sample without SHP. On the other hand, SHP may have favored the citric acid to be more involved in 278 279 crosslinkings (and less apparent as crystals), although the other results do not 280 corroborate this hypothesis. Alternatively, SHP may have favored dissolution of citric 281 acid crystals by increasing pH, or simply diluted the citric acid crystals, making them 282 less apparent.

283 **4. Conclusions**

284 Films were successfully formed using hemicelluloses extracted from wheat straw as 285 a matrix. Citric acid acted as a crosslinker, which was evidenced by its decreasing 286 effects on water solubility and water vapor permeability, and also as a plasticizer, 287 which was evident from its effects on tensile properties. Those apparently contradictory effects might be explained by a flexible crosslinking. No evidence was 288 found for the effectivity of sodium hypophosphite as catalyst for the crosslinking 289 290 reaction. FTIR spectroscopy suggested that some crosslinking reaction probably 291 took place even in a non-cured film, suggesting that the reaction may have occurred 292 even at room temperature.

293 **5. Acknowledgements**





The authors gratefully acknowledge the financial support of the UK Biotechnology & Biological Sciences Research Council (BBSRC) and the Brazilian Agricultural Research Corporation (Embrapa). Authors also thank the National Counsel of Technological and Scientific Development (CNPq, Brazil) for the Research Productivity Fellowship (304179/2012-4) granted to H.M.C. Azeredo, and for the scholarship granted to P.H.F. Pereira (243244/2013-4).

300





301 **6. References**

Abdillahi, H., Chabrat, E., Rouilly, A., & Rigal, L. (2013). Influence of citric acid on thermoplastic wheat flour/poly(lactic acid) blends. II. Barrier properties and water vapor sorption isotherms. *Industrial Crops and Products, 50,* 104-111.

ASTM (2005) Standard test methods for water vapor transmission of materials. E9605. In *Annual book of ASTM standards*. Philadelphia: American Society for Testing
and Materials.

ASTM. (2009). Standard test method for tensile properties of thin plastic sheeting.
D882-09. In *Annual Book of ASTM Standards*. Philadelphia: American Society for
Testing and Materials.

Bagheri, L., Yarmand, M., Madadlou, A., & Mousavi, M.E. (2014). Transglutaminaseinduced or citric acid-mediated cross-linking of whey proteins to tune the characteristics of subsequently desolvated sub-micron and nano-scaled particles. *Journal of Microencapsulation*, in press.

Balaguer, M.P., Gómez-Estaca, J., Gavara, R., Hernandez-Muñoz, P. (2011). Functional properties of bioplastics made from wheat gliadins modified with cinnamaldehyde. *Journal of Agricultural and Food Chemistry*, *59*, 6689-6695.

BCC Research. (2014). Global markets and technologies for bioplastics. Report
code PLS050C. Available at http://www.bccresearch.com/marketresearch/plastics/bioplastics-pls050c.html, accessed 24 July 2014.





Bonilla, J., Talón, E., Atarés, L., Vargas, M., & Chiralt, A. (2013). Effect of the incorporation of antioxidants on physicochemical and antioxidant properties of wheat starch–chitosan films. *Journal of Food Engineering*, *118*, 271-278.

Chabrat, E., Abdillahi, H., Rouilly, A., & Rigal, L. (2012). Influence of citric acid on thermoplastic wheat flour/poly(lactic acid) blends. I: Thermal, mechanical and morphological properties. *Industrial Crops and Products, 37*, 238-246.

327 Coma, V., Sebti, I., Pardon, P., Pichavant, F. H., & Deschamps, A. (2003). Film
328 properties from crosslinking of cellulosic derivatives with a polyfunctional carboxylic
329 acid. *Carbohydrate Polymers, 51,* 265–271.

330 Dastidar, T.G., & Netravali, A.N. (2012). 'Green' crosslinking of native starches with
331 malonic acid and their properties. *Carbohydrate Polymers, 90,* 1620-1628.

Fang, J.M., Sun, R.C., Salisbury, D., Fowler, P., & Tomkinson, J. (1999).
Comparative study of hemicelluloses from wheat straw by alkali and hydrogen
peroxide extractions. *Polymer Degradation and Stability*, *66*, 423-432.

Feng, X., Xiao, Z., Sui, S., Wang, Q., & Xie, Y. (2014). Esterification of wood with citric acid: The catalytic effects of sodium hypophosphite (SHP). *Holzforschung, 68,* 427-433.

Ghanbarzadeh, B., Almasi, H., & Entezami, A. A. (2011). Improving the barrier and mechanical properties of corn starch-based edible films: Effect of citric acid and carboxymethyl cellulose. *Industrial Crops And Products, 33,* 229–235.





Garcia, P.S., Grossmann, M.V.E., Shirai, M.A., Lazaretti, M.M., Yamashita, F., Muller, C.M.O., & Mali, S. (2014). Improving action of citric acid as compatibiliser in starch/polyester blown films. *Industrial Crops and Products*, *52*, 305-312.

Giancone, T., Torrieri, E., Di Pierro, P., Cavella, S., Giosafatto, C.V.L., & Masi, P. (2011). Effect of surface density on the engineering properties of high methoxyl pectin-based edible films. *Food and Bioprocess Technology*, *4*, 1228-1236.

Govumoni, S.P., Koti, S., Kothagouni, S.Y., Venkateshwar, S., & Linga, V.R. (2013).
Evaluation of pretreatment methods for enzymatic saccharification of wheat straw for
bioethanol production. *Carbohydrate Polymers*, *91*, 646-650.

Groen, H., & Roberts, K.J. (2001). Nucleation, growth, and pseudo-polymorphic behavior of citric acid as monitored in situ by attenuated total reflection Fourier Transform Infrared Spectroscopy. *The Journal of Physical Chemistry B, 105,* 10723-10730.

Hashem, M., Sharaf, S., El-Hady, M.M.A., & Hebeish, A. (2013). Synthesis and characterization of novel carboxymethylcellulose hydrogels and carboxymethylcellulolse-hydrogel-ZnO-nanocomposites. *Carbohydrate Polymers*, *95*, 421-427.

Helmut Kaiser Consultancy. (2013). Bioplastics market worldwide 2010/11-20152020-2025. Available at http://www.hkc22.com/bioplastics.html, accessed 24 July
2014.





Hu, J.-S., Song, Z.-W., Liu, C., & Zhang, W.-C. (2010). Influence of different nematic
crosslinking unit on mesomorphism of side-chain cholesteric elastomers containing
menthyl groups. *Colloid and Polymer Science*, *288*, 851-858.

Janjarasskul, T., & Krochta, J.M. (2010). Edible packaging materials. *Annual Reviews of Food Science and Technology*, *1*, 415-448.

Jia, Y.-G., Zhang, B.-Y., He, X.-Z., & Ning, J.-Z. (2005). Synthesis and phase behavior of nematic liquid-crystalline elastomers derived from smectic crosslinking agent. *Journal of Applied Polymer Science, 98*, 1712-1719.

Ma, X., Jian, R., Chang, P.R., & Yu, J. (2008). Fabrication and characterization of
citric acid-modified starch nanoparticles/plasticized starch composites. *Biomacromolecules*, *9*, 3314-3320.

Martel, F., Estrine, B., Plantier-Royon, R., Hoffmann, N., & Portella, C. (2010).
Development of agriculture left-overs: Fine organic chemicals from wheat
hemicellulose-derived pentoses. *Topics in Current Chemistry, 294,* 79–115.

Menzel, C., Olsson, E., Plivelic, T.S., Andersson, R., Johansson, C., Kuktaite, R.,
Järnstron, L., & Koch, K. (2013). Molecular structure of citric acid cross-linked starch
films. *Carbohydrate Polymers, 96,* 270-276.

378 Möller, H., Grelier, S., Pardon, P., & Coma, V. (2004). Antimicrobial and 379 physicochemical properties of chitosan-HPMC based films. *Journal of Agricultural* 380 *and Food Chemistry, 52,* 6585-6591.





Ojagh, S.M., Rezaei, M., Razavi, S.H., & Hosseini, S.M.H. (2010). Development and evaluation of a novel biodegradable film made from chitosan and cinnamon essential oil with low affinity toward water. *Food Chemistry*, *122*, 161-166.

Olivato, J.B., Grossmann, M.V.E., Bilck, A.P., & Yamashita, F. (2012). Effect of organic acids as additives on the performance of thermoplastic starch/polyester blown films. *Carbohydrate Polymers, 90*, 159-164.

Olsson, E., Hedenqvist, M.S., Johansson, C., & Järnström, L. (2013a). Influence of
citric acid and curing on moisture sorption, diffusion and permeability of starch films. *Carbohydrate Polymers, 94,* 765-772.

Olsson, E., Menzel, C., Johansson, C., Andersson, R., Koch, K., & Järnström, L.
(2013b). The effect of pH on hydrolysis, cross-linking and barrier properties of starch
barriers containing citric acid. *Carbohydrate Polymers, 98,* 1505-1513.

Peng, H., Yang, C.Q., & Wang, S. (2012). Nonformaldehyde durable press finishing
of cotton fabrics using the combination of maleic acid and sodium hypophosphite. *Carbohydrate Polymers, 87,* 491-499.

Reddy, N., & Yang, Y. (2010). Citric acid cross-linking of starch films. *Food Chemistry*, *118*, 702–711.

Reddy, N., Jiang, Q., & Yang, Y. (2012). Preparation and properties of peanut protein films crosslinked with citric acid. *Industrial Crops and Products*, *39*, 26-30.





400 Reddy, N., Li, Y., & Yang, Y. (2009). Wet cross-linking gliadin fibers with citric acid 401 and a quantitative relationship between cross-linking conditions and mechanical 402 properties. *Journal of Agricultural and Food Chemistry*, *57*, 90-98.

Rhim, J.-W., Park, H.B., Lee, C.-S., Jun, J.-H., Kim, D.S., & Lee, Y.M. (2004).
Crosslinked poly(vinyl alcohol) membranes containing sulfonic acid group: proton
and methanol transport through membranes. *Journal of Membrane Science, 238,*143-151.

Ruiz, H.A., Cerqueira, M.A., Silva, H.D., Rodríguez-Jasso, R.M., Vicente, A.A., &
Teixeira, J.A. (2013). Biorefinery valorization of autohydrolysis wheat straw
hemicellulose to be applied in a polymer-blend film. *Carbohydrate Polymers, 92,*2154-2162.

Salam, A., Pawlak, J.J., Venditti, R.A., & El-tahlawy, K. (2011). Incorporation of
carboxyl groups into xylan for improved absorbency. *Cellulose, 18,* 1033-1041.

413 Sauperl, O., & Stana-Kleinschek, K. (2010). Differences between cotton and viscose
414 fibers crosslinked with BTCA. *Textile Research Journal, 80,* 383–392.

Sebti, I., Delves-Broughton, J., & Coma, V. (2003). Physicochemical properties and
bioactivity of nisin-containing cross-linked hydroxypropylmethylcellulose films. *Journal of Agricultural and Food Chemistry*, *51*, 6468-6474.

Shi, R., Bi, J., Zhang, Z., Zhu, A., Chen, D., Zhou, X., Zhang, L., & Tian, W. (2008).
The effect of citric acid on the structural properties and cytotoxicity of the polyvinyl
alcohol/starch films when molding at high temperature. *Carbohydrate Polymers, 74,*763–770.





Shi, R., Zhang, Z., Liu, Q., Han, Y., Zhang, L., Chen, D., & Tian, W. (2007).
Characterization of citric acid/glycerol co-plasticized thermoplastic starch prepared
by melt blending. *Carbohydrate Polymers*, *69*, 748–755.

425 TAPPI. (2009). T 203 cm-99. Alpha-, beta- and gamma-cellulose in pulp.

426 Wang, S., Ren, J., Li, W., Sun, R., & Liu, S. (2014). Properties of polyvinyl 427 alcohol/xylan composite films with citric acid. *Carbohydrate Polymers, 103,* 94-99.

428 WASDE (2014). World agricultural supply and demand estimates. WASDE - 531.

429 Available at http://www.usda.gov/oce/commodity/wasde/latest.pdf, accessed 24 July430 2014.

Xiaohong, X., & Yang, C.Q. (2000). FTIR spectroscopy study of the formation of
cyclic anhydride intermediates of polycarboxylic acids catalyzed by sodium
hypophosphite. *Textile Research Journal, 70,* 64-70.

Yao, W., Wang, B., Ye, T., & Yang, Y. (2013). Durable press finishing of cotton
fabrics with citric acid: enhancement of whiteness and wrinkle recovery by polyol
extenders. *Industrial and Engineering Chemistry Research*, *52*, 16118-16127.

Yokoyama, T., Kadla, J.F., Chang, H.-M. (2002). Microanalytical method for the
characterization of fiber components and morphology of woody plants. *Journal of Agricultural and Food Chemistry, 50*, 1040–1044.

Zhang, B.-Y., Hu, J.-S., Jia, Y.-G., & Du, B.-G. (2003). Side-chain cholesteric liquid
crystalline elastomers derived from nematic bis-olefinic crosslinking units. *Macromolecular Chemistry and Physics*, *204*, 2123-2129.







- 1 **Figure 1.** Proposed mechanism for covalent crosslinking between citric acid and a
- 2 polysaccharide. (Adapted from Hashem et al., 2013).
- **Figure 2.** Film properties as affected by citric acid (CA) concentration and by the presence of sodium hypophosphite. CA-C: films added with citric acid; CA-C-SHP: films added with citric acid and sodium hypophosphite; CA-NC: films added with citric acid, but not subjected to the curing treatment. Error bars represent standard error of the mean. Values in the same line labelled with the same letter are not significantly different (Tukey, p<0.05).
- 9 Figure 3. FTIR spectroscopy of films: CA0 (control), CA30-C (30% citric acid, cured),
- 10 CA30-C-SHP (30% citric acid + 15% SHP, cured), and CA30-NC (30% citric acid,
- 11 non-cured).
- Figure 4. SEM images of films with 30% citric acid without SHP (A) or with SHP (B),
 and the control (C).

download high resolution image

Fig





































Highlights

Hemicelluloses were extracted from wheat straw to produce films.

Hemicelluloses were added with citric acid, with or without sodium hypophosphite.

Citric acid acted both as a crosslinker and a plasticizer to hemicellulose films.

Sodium hypophosphite did not improve film properties.

Some crosslinking reaction occurred even when the films were not cured at 150°C.





Table 1. Holocellulose, α -cellulose and hemicellulose contents (on a dry weight basis) of the wheat straw and the hemicellulose powder (values represented as mean ± standard deviation).

Fractions	Milled wheat straw	Hemicellulose powder
Holocellulose (%)	62.52 ± 0.86	91.21 ± 0.74
α-cellulose (%)	$\textbf{32.62} \pm \textbf{1.63}$	$\textbf{4.14} \pm \textbf{0.17}$
Hemicelluloses (%)	29.90 ± 0.96	87.06 ± 0.79